

RESEARCH ARTICLE

Ecological quality of the Sea of Marmara (Turkey) assessed by the Marine Floristic Ecological Index (MARFEI)

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Abstract

Benthic macrophytes (macroalgae and angiosperms) are proposed as biological elements to assess the ecological status of coastal and transitional waters within the EU Water Framework Directive. In the present paper, a sampling of benthic macrophytes was made from 0-5 m depth at 25 different sites of the Sea of Marmara (Turkey) in 2015 and the *Marine Floristic Ecological Index* (MARFEI) was applied. Macrophytes were classified into five ecological status groups: ESGI (IA, IB, IC; late-successional taxa such as *Cystoseira* spp., angiosperms, calcareous algae) and ESGII (IIA, IIB; opportunistic taxa such as filamentous and sheet-like green algae). The study revealed good ecological status class (ESC) for 1 site, moderate for 11 sites, poor for 10 sites, and bad for 3 sites. MARFEI_{EQR} showed a strong negative linear relationship with the pressure index MA-LUSI. Nevertheless, the results of this study have to be validated by similar studies in other geographical areas.

Keywords: Angiosperms, MARFEI, pressures, marine algae, taxonomy

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Introduction

The EU Water Framework Directive (WFD, 2000/60/EC) requires the establishment of monitoring programs to assess the ecological status in coastal and transitional waters throughout Europe. To facilitate this task, macroalgae and

angiosperms were proposed as key biological quality elements. Marine benthic macrophytes, especially seagrasses and perennial macroalgae, are sensitive to anthropogenic activities, especially those linked with nutrient input and degradation of light regime in the water leading to eutrophication. For this reason several macrophyte indices have been proposed for the Mediterranean eco-region. These indices include the following: the Ecological Evaluation Index (EEI) (Orfanidis *et al.* 2001, 2011), the *Posidonia oceanica* multivariate index (POMI) (Romero *et al.* 2007), the CARtography of LITtoral (CARLIT) (Ballesteros *et al.* 2007), the Macrophyte Quality Index (MaQI) (Sfriso *et al.* 2007, 2009), the *Posidonia oceanica* Rapid Easy Index (PREI) (Gobert *et al.* 2009), and the Alien Biotic Index (ALEX) (Çınar and Bakır 2014; Piazzzi *et al.* 2015). Recently, EEI and CARLIT for coastal waters, and EEI and MAQI for transitional waters have been successfully intercalibrated by the Mediterranean Geographic Intercalibration Group (MEDGIG) (EC 2013), to ensure that good ecological status represents the same level ecological quality across the Mediterranean eco-region.

EEI has been applied in Greece, Cyprus, and Slovenia (Orfanidis *et al.* 2001, 2011; Panayotidis *et al.* 2004; Orlando-Bonaca *et al.* 2008). Nikolić *et al.* (2011) tested both CARLIT and EEI to measure the ecological status of the Adriatic coast of Croatia. Macrophyte biotic indices (EEI, CARLIT, and MaQI) were tested for the ecological quality classification of Turkish coastal and transitional waters by Taşkın and Öztürk (2014a, b), Taşkın *et al.* (2013, 2016), Dağlar and Taşkın (2015), Taşkın (2015).

In the present study, a new method, namely, the *Marine Floristic Ecological Index* (MARFEI), was tested to assess the ecological status classes at 25 sites of the Sea of Marmara (Turkey). The concept of the MARFEI index is similar to EEI, and it is based on the obvious and universal pattern that “anthropogenic disturbance, e.g. pollution, eutrophication, shifts the ecosystem from pristine to more degraded states, and where opportunistic species are dominant” (Orfanidis *et al.* 2001). The relationship between the pressure index MA-LUSI and MARFEI_{EQR} is also tested.

Materials and Methods

Study area

The Sea of Marmara is bordered completely by Turkey. It connects the Black Sea to the Aegean Sea, and it is connected to the Black Sea by the Istanbul (Bosphorus) Strait and to the Aegean Sea by the Çanakkale (Dardanelles) Strait. There are many islands (the Marmara Island, Paşalimanı Island, Princes Islands, etc.) in the Sea of Marmara. Sampling was made at 25 localities in the Sea of Marmara (Turkey) in 2015 (Figure 1).

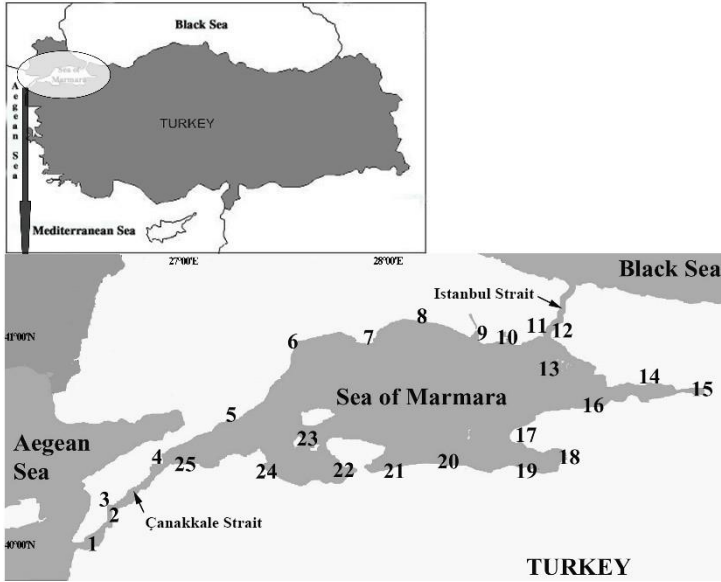


Figure 1. Research area and sampling stations in the Sea of Marmara (Turkey) [1-İntepe; 2-Çanakkale; 3-Eceabat; 4-Gelibolu; 5-Şarköy; 6-Tekirdağ; 7-M.Ereğlisi; 8-Silivri; 9-B.Çekmece; 10-K.Çekmece; 11-Haliç; 12-Üsküdar; 13-Princes Islands; 14-Hereke; 15-Kocaeli; 16-Yalova; 17-Armutlu; 18-Gemlik; 19-Mudanya; 20-Susurluk River-Boğaz; 21-Bandırma; 22-Erdek; 23-Paşalimanı Island; 24-Karabiga; 25-Lapseki]

Sampling

The material was collected by snorkeling from a 100 x100 m area per station in May and November 2015, and specimens were preserved in 2-5% formaldehyde in sea water. Samples were studied using a light microscope (Nikon SE), and voucher specimens are deposited in the personal herbaria of Ergün Taşkın (ET) in the Department of Biology of the Manisa Celal Bayar University (Manisa, Turkey). Nomenclature was checked against Guiry and Guiry (2018).

Environmental parameters

Physico-chemical parameters (salinity, temperature, pH, conductivity, turbidity, and dissolved oxygen) were measured by Water Quality Checker™ (DKK-TOA WQC 24). Ortho-phosphate (Parsons *et al.* 1984), and ammonium nitrogen (Strickland and Parsons, 1972) were also measured (Table 1).

Table 1. The average physico-chemical parameters of the sampling stations (T: temperature, DO:dissolved oxygen; Turb.: turbidity; Cond.: conductivity; Sal.: salinity; P: phosphate; AN: ammonium nitrogen)

Stations	pH	T(°C)	DO (mg/L)	Turb. (NTU)	Cond. (S/m)	Sal. (‰)	P (mg/L)	AN (mg/L)
İtepe	8.42	20.1	8.25	0	3.78	23.8	0.325	0.045
Çanakkale	8.30	16	6.7	0	3.68	22.9	0.297	0.049
Eceabat	8.41	16.3	8.6	0	3.68	23.3	0.322	0.045
Gelibolu	8.16	15.41	6.09	0	3.56	21.9	0.289	0.048
Şarköy	8.44	15.1	9.75	0	3.54	21.8	0.259	0.046
Tekirdağ	8.26	18.3	7.56	2	3.29	20.4	0.298	0.054
Marmara Ereğlisi	8.29	23.2	7.3	10.2	3.14	19.5	0.304	0.045
Silivri	8.20	23.6	7.5	19.6	2.48	15.1	0.311	0.049
Büyükçekmece	8.50	19.4	10.1	11.9	2.98	18.3	0.289	0.042
Küçükçekmece	8.82	18.9	10.26	13.6	2.16	13.1	0.262	0.041
Haliç	7.91	14.9	5.93	0.9	2.56	15.2	0.274	0.05
Üsküdar	8.17	12.4	7.58	0	3.03	17.8	0.237	0.048
Princes Islands	8.38	17.2	9.32	3.7	3.19	19.4	0.249	0.041
Hereke	8.32	18.5	4.81	5.2	3.15	19.3	0.251	0.043
Kocaeli	8.65	18.9	9.56	1.4	3.51	21.9	0.249	0.042
Yalova	8.30	17.1	7.65	0	3.52	22	0.292	0.046
Armutlu	8.59	21.8	9.5	12.1	3.39	21.3	0.263	0.038
Gemlik	8.32	23	6.56	37	2.12	12.7	0.264	2.161
Mudanya	8.54	20.5	8.86	1.1	2.83	17.3	0.308	0.042
Susurluk River-Boğaz	8.67	22.7	11.45	2.3	3.05	18.9	0.287	0.049
Bandırma	8.39	21	8.37	5	3.46	21.6	0.289	0.051
Erdek	8.38	18.1	8.43	0.7	3.83	24.1	0.285	0.052
Paşalimanı Island	8.35	17.6	8.05	0	3.5	21.6	0.333	0.055
Karabiga	8.54	23.3	10.35	2.7	3.59	22.7	0.31	0.051
Lapseki	8.48	19.7	9.2	2.7	3.65	22.8	0.156	0.059

Pressure data

The Macroalgae-Land Uses Simplified Index (MA-LUSI) has been developed as an improvement of LUSI index (Flo *et al.* 2011) using pressures affecting the benthic macrophytes. The MA-LUSI index takes under account the pressures from mariculture, sewage outfall, harbours, irregular fresh water inputs, sediment nutrient release, urban, commercial and industrial, agriculture, and background trophic status affect a 3 km buffer zone around the sampling site. It has been developed by the MEDGIG of macroalgae (EC 2013).

Data analysis

Macrophytes (macroalgae and angiosperms) were classified into five ecological status groups, ESGI (IA, IB, IC; late-successional taxa such as *Cystoseira* spp., angiosperms, calcareous algae), and ESGII (IIA, IIB; opportunistic taxa such as filamentous and sheet-like green algae) (Orfanidis *et al.* 2011) (Appendix 1). Ecological quality ratio (EQR) between 0 and 1 was obtained by the formulation (bad=0-0.20; poor=0.20-0.40; moderate=0.40-0.60; good=0.60-0.80; high=0.80-1).

Formula;

$$\text{MARFEI}_{\text{EQR}} = (4 * \%IA + 3 * \%IB + 2 * \%IC) / (1 * IIA + 0,5 * IIB)$$

The relationship between pressures and $\text{MARFEI}_{\text{EQR}}$ values was calculated using the MA-LUSI index.

Results and Discussion

Physicochemical parameters were found as pH min. 7.91 – max. 8.82, temperature (°C) min. 12.4 – max. 23.6, salinity min. 12.7 – max. 23.8, dissolved oxygen (mg/L) min. 4.81 – max. 11.45, turbidity (NTU) min. 0 – max. 37, conductivity (S/m) min. 2.12 – max. 3.78, phosphate (mg/L) min. 0.156 – max. 0.333, and ammonium nitrogen (mg/L) min. 0.038 – max. 2.161 in the Sea of Marmara. The lowest temperature was measured in Üsküdar (12.4 °C) because of Bosphorus streams from the Black Sea to the Aegean Sea via the Sea of Marmara. The highest value of ammonium nitrogen was measured in Gemlik (2.161 mg/L).

The Çanakkale Strait is situated between the Aegean Sea and the Sea of Marmara and has sandy, stony and rocky bottoms. Five stations (İntepe, Çanakkale, Eceabat, Gelibolu, and Lapseki) were selected on the coast of the Strait characterized by salinity between 21.9‰ and 23.8‰. *Cystoseira* spp., *Padina pavonica*, calcareous red algae, and the marine angiosperms *Posidonia oceanica* and *Cymodocea nodosa* were abundant there. Şarköy station was located in the Sea of Marmara and was characterized by sandy and partially by stony substratum covered by *Cystoseira barbata*, *Cymodocea nodosa*, calcareous red algae, and the green algae *Ulva* spp. and *Cladophora* spp. Tekirdağ and Marmara Ereğlisi had sandy and stony substratum, and these stations have been affected by harbours. Silivri, Büyükçekmece, Küçükçekmece, Haliç, and Üsküdar were all impacted by the antropogenic activities of the city of İstanbul, and opportunistic algae (*Ulva* spp., *Cladophora* spp., *Ceramium* spp., and blue-green algae) are found as dominant. The calcareous red algae *Corallina officinalis* and *Ellisolandia elongata* were abundant in Üsküdar station, where it is effected by the Bosphorus stream. Princes Islands (Adalar) consist of nine Islands (Büyükkada, Heybeliada, Burgazada, Kınalıada, Sedef Adası, Yassıada, Sivriada, Kaşık Adası), and Tavşan

Adası (Neandros Island), and sampling was mainly made in Büyükada. Hereke and Kocaeli have a sandy and partially stony substratum. This area is impacted by commercial and industrial activities, and opportunistic algae (*Ulva rigida*, *Gracilaria gracilis*, etc.) were dominant. Yalova and Armutlu stations were characterized by sandy and stony substrata, and both sensitive (*Cystoseira barbata*, *Cymodocea nodosa*, calcareous red algae) and opportunistic algae (*Ulva* spp. and *Cladophora* spp.) were present there. Gemlik was very much impacted by commercial, industrial activities, and urbanization, and *Ulva rigida*, *Ceramium virgatum*, and blue-green algae were dominant. Mudanya has a stony and sandy substratum, and *Cystoseira barbata* and calcareous red algae were common. Susurluk River-Boğaz was characterized by sandy and partially by stony substratum, and it is affected by Susurluk River, and opportunistic algae dominated. Bandırma was impacted by urbanization as well as by harbour and shipping pressures, and that was characterized by blooms of opportunistic algae. Paşalimanı Island was characterized by sandy and by stony substratum, and both sensitive and opportunistic algae were found in abundance. Erdek and Karabiga stations were impacted by agriculture; they had a stony and sandy substratum. The alien and invasive green alga *Codium fragile* subsp. *fragile* was generally common in all stations of the Sea of Marmara from coast to depths of 20 m.

A total of 276 seaweed and marine angiosperm taxa [89 Phaeophyceae (brown algae), 139 Rhodophyta (red algae), 45 Chlorophyta (green algae), and three Spermatophyta] at specific and infraspecific levels are reported from 25 different localities in the Sea of Marmara (Appendix 1). The highest number of taxa was found in Paşalimanı Island (157 taxa), and Şarköy (154 taxa), İntepe (152 taxa), and Gelibolu (134), while the lowest number of taxa was found in Kocaeli (28 taxa), Hereke (30 taxa), Susurluk River-Boğaz (31 taxa), Gemlik (32 taxa), and Silivri (36 taxa), respectively.

Macrophytes (macroalgae and angiosperms) are known as good bio-indicators and as biological elements to assess the ecological status of coastal and transitional waters according to WFD. The Marine Floristic Ecological Index (MARFEI) used the number of taxa of five ESG's (ESG IA, IB, IC, and ESG IIA, IIB) (Orfanidis *et al.* 2011) in the Sea of Marmara. The Sea of Marmara is high marine traffic area moderately affected by anthropogenic disturbances except in locations near urban or industrial centers such as İstanbul. Indeed, MARFEI classified the Marmara Sea as various status, 1 site (İntepe) was classified as good ESC, 11 sites (Eceabat, Şarköy, Marmara Ereğlisi, Üsküdar, Yalova, Armutlu, Mudanya, Erdek, Paşalimanı Island, Karabiga, and Lapseki) as moderate, 10 sites (Çanakkale, Gelibolu, Tekirdağ, Silivri, Küçükçekmece, Büyükçekmece, Gemlik, Susurluk River-Boğaz, Princes Islands, and Bandırma) as poor, and 3 sites (Kocaeli, Hereke, and Haliç) as bad ESC (Table 2 and Figure 2). The relationship between the pressure MA-LUSI index and MARFEI_{EQ}R was tested, and a negative linear relationship was found (Figure 3). In Kocaeli, Haliç, and Gemlik sites that were under the influence of heavy pollution due to existence of urban, commercial

and industrial sewage outfall, and harbours, MA-LUSI values were maximum, and the green algae *Ulva* spp. and *Cladophora* spp., and the red algae *Ceramium* spp. and *Gracilaria gracilis* were at highest abundance. On the other side, in the less impacted, pristine sites of İntepe, Lapseki, Mudanya, Şarköy, Armutlu, Erdek, Eceabat, and Paşalimanı Island, MA-LUSI values were found minimum and the late-successional, sensitive macroalgae (*Cystoseira* spp., *Padina pavonica*, calcareous algae, etc.) and marine angiosperms (*Posidonia oceanica*, *Cymodocea nodosa*) were most abundant.

The macrophyte biotic indices EEI, CARLIT, MaQI have been applied in several studies to assess the ESC's of Turkish coastal and transitional waters (Taşkın and Öztürk 2014a,b; Taşkın *et al.* 2013, 2016; Dağlar and Taşkın 2013, 2015; Taşkın 2015). MARFEI is supplementary to EEI index designed to assess the ESC in sites where only presence-absence benthic macrophyte data are available giving the possibility also to use in water quality assessment the old check-lists available within the Mediterranean Sea.

In conclusion, MARFEI after its validation in other geographical areas could be recommended for future monitoring activities for Turkish coastal and transitional waters having as a prerequisite a thorough knowledge of benthic macrophyte biodiversity.

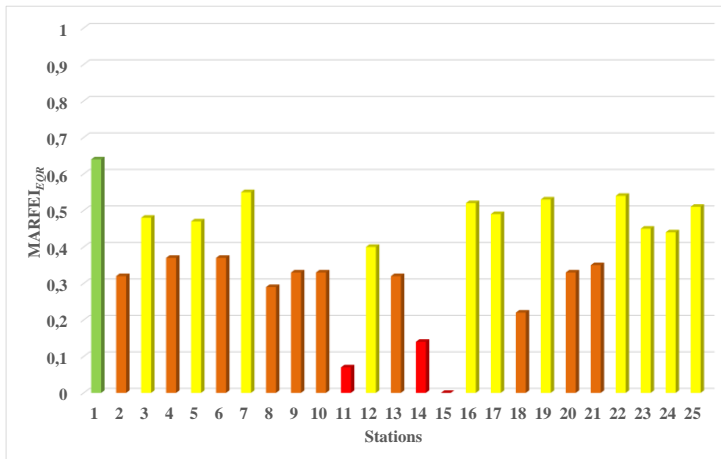


Figure 2. The Ecological Quality Ratio of station in the Sea of Marmara (Turkey) by MARFEI_{EQR}. [1-İntepe; 2-Çanakkale; 3-Eceabat; 4-Gelibolu; 5-Şarköy; 6-Tekirdağ; 7-M.Ereğlisi; 8-Silivri; 9-B.Çekmece; 10-K.Çekmece; 11-Haliç; 12-Üsküdar; 13-Princes Islands; 14-Hereke; 15-Kocaeli; 16-Yalova; 17-Armutlu; 18-Gemlik; 19-Mudanya; 20-Susurluk River-Boğaz; 21-Bandırma; 22-Erdek; 23-Paşalimanı Island; 24-Karabiğa; 25-Lapseki]

Table 2. The Ecological Quality Ratio by MARFEI_{EQR} and pressures data (MA-LUSI index) in the Sea of Marmara (Turkey)

Stations	MARFEI _{eqr}	MA-LUSI	Species richnees
İntepe	0,64	2	152
Çanakkale	0,32	5	87
Eceabat	0,48	3	131
Gelibolu	0,37	4	134
Şarköy	0,47	3	154
Tekirdağ	0,37	7	68
Marmara Ereğlisi	0,55	5	76
Silivri	0,29	7,8125	36
Büyükçekmece	0,33	6,25	41
Küçükçekmece	0,33	7,8125	54
Haliç	0,07	12,5	39
Üsküdar	0,4	6,25	55
Princes Islands	0,32	2,8125	125
Hereke	0,14	8,75	30
Kocaeli	0	10,9375	28
Yalova	0,52	5	70
Armutlu	0,49	3	71
Gemlik	0,22	12,5	32
Mudanya	0,53	4	67
Susurluk River-Boğaz	0,33	3,75	31
Bandırma	0,35	5	75
Erdek	0,54	4	84
Paşalimanı Island	0,45	2	157
Karabiga	0,44	3,75	73
Lapseki	0,51	2	127

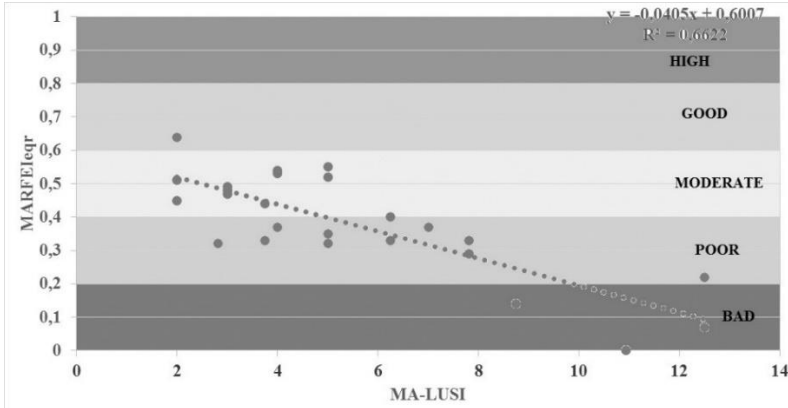


Figure 3. Relation between MARFEI_{EQR} and pressure data (MA-LUSI index) in the Sea of Marmara (Turkey)

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Denizel Floristik Ekolojik İndeks (DENFEİ) Metodu ile Marmara Denizi'nin (Türkiye) ekolojik durumunun belirlenmesi

Öz

AB Su Çerçeve Direktifi'nde kıyı ve geçiş sularının ekolojik durumunun değerlendirilmesi için bentik makrofitler (makroalgler ve angiospermler) biyolojik element olarak önerilmektedir. Bu çalışmada, 2015 yılında Marmara Denizi'nin (Türkiye) 25 farklı noktasından 0-5 m derinlikten bentik makrofitler örnekledi ve Denizel Floristik Ekolojik İndeks (DENFEİ) uygulanmıştır. Makrofitler, ESGI (IA, IB, IC; *Cystoseira* spp., angiospermler, kalkerli algler gibi geç gelişimsel gösteren taksonlar) ve ESGII (IIA, IIB; ipliksi ve yapraklı yeşil algler gibi fırsatçı taksonlar). Bu çalışmada, 1 nokta için iyi, 11 nokta için orta, 10 nokta için zayıf ve 3 nokta için kötü ekolojik durum sınıfında (EDS) tespit edilmiştir. DENFEİ_{EKO}, baskı indeksi MA-LUSI ile kuvvetli negatif bir doğrusal ilişki göstermiştir. Bu çalışmanın sonuçları diğer coğrafik bölgelerde benzer çalışmalarla da doğrulanmalıdır.

Anahtar kelimeler: Angiospermler, baskılar, deniz algleri, DENFEI, taksonomi

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Appendix 1. The Ecological State Groups (ESG*) of marine benthic macrophytes (macroalgae and angiosperms)
(IA, IB, IC=sensitive taxa; IIA, IIB=tolerant-opportunist taxa).

Taxa	ESG
MACROALGAE	
<i>Acetabularia acetabulum</i> (L.) P.C.Silva	IC
<i>Acinetospora crinita</i> (Carmichael) Sauvageau	IIB
<i>Acrochaetium crassipes</i> (Børgesen) Børgesen	IIB
<i>Acrochaetium mahumetanum</i> Hamel	IIB
<i>Acrochaetium microscopicum</i> (Nägeli ex Kützing) Nägeli	IIB
<i>Acrochaetium parvulum</i> (Kylin) Hoyt	IIB
<i>Acrochaetium savianum</i> (Meneghini) Nägeli	IIB
<i>Acrochaetium secundatum</i> (Lyngbye) Nägeli	IIB
<i>Acrochaetium virgatulum</i> (Harvey) Batters	IIB
<i>Acrodiscus vidovichii</i> (Meneghini) Zanardini	IIA
<i>Acrosorium ciliolatum</i> (Harvey) Kylin	IIB
<i>Aglaothamnion</i> sp.	IIB
<i>Aglaothamnion tenuissimum</i> (Bonnemaison) Feldmann-Mazoyer	IIB
<i>Aglaothamnion tripinnatum</i> (C.Agardh) Feldmann-Mazoyer	IIB
<i>Alsidium corallinum</i> C.Agardh	IIA
<i>Amphiroa rigida</i> J.V. Lamouroux	IC
<i>Anotrichium furcellatum</i> (J.Agardh) Baldock	IIB
<i>Anotrichium tenue</i> (C.Agardh) Nägeli	IIB
<i>Antithamnion cruciatum</i> (C.Agardh) Nägeli	IIB
<i>Antithamnion hubsii</i> E.Y.Dawson	IIB
<i>Arthrocladia villosa</i> (Hudson) Duby	IIA
<i>Asperococcus bullosus</i> J.V.Lamouroux	IIA
<i>Asperococcus ensiformis</i> (Della Chiaje) M.J. Wynne	IIA
<i>Asperococcus fistulosus</i> (Hudson) W.J.Hooker	IIA
<i>Bangia atropurpurea</i> (Mertens ex Roth) C.Agardh	IIB
<i>Blidingia marginata</i> (J.Agardh) P.Dangeard ex Bliding	IIB
<i>Boergeseniella fruticulosa</i> (Wulfen) Kylin	IIB
<i>Boergeseniella thuyoides</i> (Harvey) Kylin	IIB
<i>Botrytella micromora</i> Bory de Saint-Vincent	IIB
<i>Botrytella parva</i> (Takamatsu) H.-S. Kim	IIB
<i>Bryopsis corymbosa</i> J.Agardh	IIB
<i>Bryopsis hypnoides</i> J.V.Lamouroux	IIB
<i>Bryopsis plumosa</i> (Hudson) C.Agardh	IIB
<i>Callithamnion corymbosum</i> (Smith) Lyngbye	IIB
<i>Caulerpa cylindracea</i> Sonder	IIB
<i>Ceramium ciliatum</i> var. <i>robustum</i> (J.Agardh) Mazoyer	IIB
<i>Ceramium cimbricum</i> H.E.Petersen	IIB
<i>Ceramium circinatum</i> (Kützing) J.Agardh	IIB
<i>Ceramium deslongchampsii</i> Chauvin ex Duby	IIB
<i>Ceramium diaphanum</i> (Lightfoot) Roth	IIB
<i>Ceramium secundatum</i> Lyngbye	IIB
<i>Ceramium siliquosum</i> f. <i>minusculum</i> (Feldmann-Mazoyer) Gómez Garreta, T.Gallardo, M.Ribera, Cormaci, G.Furnari, G. Giaccone & C.F.Boudour.	IIB
<i>Ceramium siliquosum</i> var. <i>elegans</i> (Roth) G. Furnari	IIB
<i>Ceramium siliquosum</i> var. <i>lophophorum</i> (Feldmann-Mazoyer) Serio	IIB
<i>Ceramium tenerrimum</i> (G.Martens) Okamura	IIB
<i>Ceramium tenerrimum</i> var. <i>brevizonatum</i> (H.E. Petersen) Feldmann-Mazoyer	IIB
<i>Ceramium virgatum</i> Roth	IIB
<i>Ceramium virgatum</i> var. <i>implexo-contortum</i> (Solier) G.Furnari	IIB
<i>Chaetomorpha ligustica</i> (Kützing) Kützing	IIB

Appendix 1. Continued

<i>Chaetomorpha linum</i> (O.F.Müller) Kützing	IIB
<i>Champia parvula</i> (C.Agardh) Harvey	IIA
<i>Chondracanthus acicularis</i> (Roth) Fredericq	IIA
<i>Chondria capillaris</i> (Hudson) M.J. Wynne	IIA
<i>Chondria dasyphylla</i> (Woodward) C.Agardh	IIA
<i>Chondria mairei</i> G.Feldmann	IIA
<i>Chroodactylon ornatum</i> (C. Agardh) Basson	IIB
<i>Chylocladia verticillata</i> (Lightfoot) Bliding	IIA
<i>Cladophora albida</i> (Nees) Kützing	IIB
<i>Cladophora coelothrix</i> Kützing	IIB
<i>Cladophora dalmatica</i> Kützing	IIB
<i>Cladophora flexuosa</i> (O.F.Müller) Kützing	IIB
<i>Cladophora glomerata</i> (L.) Kützing	IIB
<i>Cladophora hutchinsiae</i> (Dillwyn) Kützing	IIB
<i>Cladophora laetevirens</i> (Dillwyn) Kützing	IIB
<i>Cladophora lehmanniana</i> (Lindenberg) Kützing	IIB
<i>Cladophora nigrescens</i> Zanardini	IIB
<i>Cladophora pellucida</i> (Hudson) Kützing	IIB
<i>Cladophora prolifera</i> (Roth) Kützing	IIB
<i>Cladophora rupestris</i> (L.) Kützing	IIB
<i>Cladophora sericea</i> (Hudson) Kützing	IIB
<i>Cladophora vagabunda</i> (L.) C.Hoek	IIB
<i>Cladosiphon contortus</i> (Thuret) Kylin	IIB
<i>Cladosiphon mediterraneus</i> Kützing	IIB
<i>Cladosiphon zosterae</i> (J. Agardh) Kylin	IIB
<i>Cladostephus spongiosus</i> (Hudson) C.Agardh	IIA
<i>Codium fragile</i> subsp. <i>fragile</i> (Suringar) Hariot	IIB
<i>Codium tomentosum</i> Stackhouse	IIB
<i>Codium vermilara</i> (Olivi) Delle Chiaje	IIB
<i>Colaconema codicola</i> (Børgesen) H.Stegenga, J.J.Bolton, & R.J.Anderson	IIB
<i>Colaconema daviesii</i> (Dillwyn) Stegenga	IIB
<i>Colpomenia peregrina</i> Sauvageau	IIA
<i>Colpomenia sinuosa</i> (Mertens ex Roth) Derbès et Solier	IIA
<i>Compsothamnion thuyoides</i> (Smith) Nägeli	IIB
<i>Corallina officinalis</i> L.	IC
<i>Corynophlaea umbellata</i> (C. Agardh) Kützing	IIB
<i>Cutleria chilosa</i> (Falkenberg) P.C. Silva	IB
<i>Cystoseira amentacea</i> var. <i>stricta</i> Montagne	IA
<i>Cystoseira barbata</i> (Stackhouse) C.Agardh	IB
<i>Cystoseira bosporica</i> Sauvageau	IA
<i>Cystoseira compressa</i> (Esper) Gerloff et Nizamuddin	IB
<i>Cystoseira crinita</i> Duby	IA
<i>Cystoseira foeniculacea</i> (L.) Greville	IA
<i>Cystoseira foeniculacea</i> f. <i>tenuiramosa</i> (Ercegovic) A.Gómez Garreta, M.C. Barceló, M.A. Ribera & J.Rull Lluçh	IA
<i>Dasya baillouviana</i> (S.G.Gmelin) Montagne	IIB
<i>Dasya corymbifera</i> J.Agardh	IIB
<i>Dasya hutchinsiae</i> Harvey	IIB
<i>Dasya rigidula</i> (Kützing) Ardissonne	IIB
<i>Dermocorynus dichotomus</i> (J.Agardh) Gargiulo, M.Morabito & Manghisi	IIA
<i>Dictyota dichotoma</i> (Hudson) J.V.Lamouroux var. <i>dichotoma</i>	IIA
<i>Dictyota dichotoma</i> var. <i>intricata</i> (C.Agardh) Greville	IIA
<i>Dictyota fasciola</i> (Roth) J.V.Lamouroux	IIA
<i>Dictyota linearis</i> (C.Agardh) Greville	IIA

Appendix 1. Continued

<i>Dictyota spiralis</i> Montagne	IIA
<i>Ectocarpus crouaniorum</i> Thuret	IIB
<i>Ectocarpus fasciculatus</i> Harvey	IIB
<i>Ectocarpus siliculosus</i> (Dillwyn) Lyngbye	IIB
<i>Ectocarpus siliculosus</i> var. <i>dasycaucus</i> (Kuckuck) Gallardo	IIB
<i>Ectocarpus siliculosus</i> var. <i>hiemalis</i> (P.L. et H.M. Crouan ex Kjellman) Gallardo	IIB
<i>Ectocarpus siliculosus</i> var. <i>penicillatus</i> C. Agardh	IIB
<i>Ectocarpus</i> sp.	IIB
<i>Elachista stellaris</i> J.E. Areschoug	IIB
<i>Ellisolandia elongata</i> (J.Ellis & Sol.) K.R.Hind & Saunders	IC
<i>Erythrotrichia</i> sp.	IIB
<i>Erythrotrichia carnea</i> (Dillwyn) J.Agardh	IIB
<i>Eudesme virescens</i> (Carmichael ex Berkeley) J. Agardh	IIB
<i>Falkenbergia rufolonosa</i> (Harv.) F. Schmitz Tetrasporophyte of <i>Asparagopsis armata</i> Harv.	IIB
<i>Feldmannia globifera</i> (Kützing) Hamel	IIB
<i>Feldmannia irregularis</i> (Kützing) G. Hamel	IIB
<i>Feldmannia lebelii</i> (P.L. et H.M. Crouan) G. Hamel	IIB
<i>Feldmannia mitchelliae</i> (Harv.) H.-S.Kim	IIB
<i>Feldmannia padinae</i> (Buffham) G. Hamel	IIB
<i>Feldmannia paradoxa</i> (Montagne) G. Hamel	IIB
<i>Feldmannia paradoxa</i> var. <i>donatae</i> (Ercegovic) Antolic & Span	IIB
<i>Ganonema farinosum</i> (J.V. Lamouroux) K.C. Fan et Yung C. Wang	IC
<i>Gayliella flaccida</i> (Harvey ex Kützing) T.O.Cho & L.J. McIvor	IIB
<i>Gayliella mazoyerae</i> T.O.Cho, Fredericq & Hommersand	IIB
<i>Gayralia oxysperma</i> (Kützing) K.L.Vinogradova ex Scagel et al.	IIB
<i>Gelidiella nigrescens</i> (Feldmann) Feldmann & G.Hamel	IIA
<i>Gelidium crinale</i> (Hare ex Turner) Gaillon	IIA
<i>Gelidium pusillum</i> (Stackh.) Le Jolis	IIA
<i>Gelidium serra</i> (S.G.Gmelin) E.Taşkın & M.J. Wynne	IIA
<i>Gelidium spinosum</i> (S.G.Gmelin) P.C.Silva	IIA
<i>Giraudia sphacelarioides</i> Derbès et Solier	IIB
<i>Gracilaria bursa-pastoris</i> (S.G.Gmelin) P.C.Silva	IIA
<i>Gracilaria gracilis</i> (Stackh.) M.Steentoft, L.M. Irvine & W.F.Farnham	IIA
<i>Gymnogongrus griffithsiae</i> (Turner) Martius	IIA
<i>Haliptilon attenuatum</i> (Kützing) Garbary et Johansen	IC
<i>Haliptilon virgatum</i> (Zanardini) Garbary & H.W.Johansen	IC
<i>Halopithys incurva</i> (Hudson) Batters	IB
<i>Halopteris filicina</i> (Grateloup) Kützing	IIA
<i>Halopteris scoparia</i> (L.) Sauvageau	IIA
<i>Halothrix lumbricalis</i> (Kützing) Reinke	IIB
<i>Halurus flosculosus</i> (J. Ellis) Maggs & Hommers.	IIB
<i>Halymenia latifolia</i> P.Crouan & H.Crouan ex Kützing	IIA
<i>Halymenia</i> sp.	IIA
<i>Hecatonema terminale</i> (Kützing) Kylin	IIB
<i>Herposiphonia secunda</i> (C. Agardh) Ambronn	IIB
<i>Herposiphonia tenella</i> (C.Agardh) Ambronn	IIB
<i>Heterosiphonia crispella</i> (C. Agardh) M.J. Wynne	IIB
<i>Hincksia fuscata</i> (Zanardini) P.C. Silva	IIA
<i>Hincksia granulosa</i> (J.E. Smith) P.C. Silva	IIB
<i>Hincksia sandriana</i> (Zanardini) P.C. Silva	IIB
<i>Hydroolithon farinosum</i> (J.V.Lamouroux) D. Penrose et Y.M. Chamberlain	IC
<i>Hypnea musciformis</i> (Wulfen) J.V. Lamouroux	IIA
<i>Hypoglossum hypoglossoides</i> (Stackhouse) F.Collins et Hervey	IIA

Appendix 1. Continued

<i>Jania adhaerens</i> J.V.Lamouroux	IC
<i>Jania rubens</i> (L.) J.V.Lamouroux	IC
<i>Kallymenia requienii</i> (J.Agardh) J.Agardh	IIA
<i>Kuckuckia spinosa</i> (Kützing) Kornmann	IIB
<i>Kützingiella battersii</i> (Bornet) Kornmann	IIB
<i>Laurencia obtusa</i> (Huds.) J.V.Lamouroux	IIA
<i>Laurencia pyramidalis</i> Bory ex Kützing	IIA
<i>Laurencia uvifera</i> (Forssk.) Børgesen	IIA
<i>Lithophyllum corallinae</i> (P.L. et H.M.Crouan) Heydrich	IC
<i>Lithophyllum cystoseirae</i> (Hauck) Heydrich	IC
<i>Lomentaria clavellosa</i> (Lighfoot ex Turner) Gaillon	IIA
<i>Lomentaria clavellosa</i> f. <i>urvillei</i> (J.V.Lamouroux) E.Taşkın	IIA
<i>Lomentaria ercegovicii</i> Verlaque, Boudour., Meinesz, Giraud & Marcot Coqueugniot	IIA
<i>Lophosiphonia cristata</i> Falkenberg	IIB
<i>Lophosiphonia obscura</i> (C. Agardh) Falkenberg	IIB
<i>Melobesia membranacea</i> (Esper) J.V. Lamouroux	IC
<i>Meredithia microphylla</i> (J. Agardh) J. Agardh	IIA
<i>Mesogloia lanosa</i> P.L. et H.M. Crouan	IIB
<i>Mesogloia leveillei</i> (J. Agardh) Meneghini	IIB
<i>Mesogloia vermiculata</i> (J.E. Smith) S.F. Gray	IIB
<i>Mesophyllum lichenoides</i> (J. Ellis) M. Lemoine	IC
<i>Microcoryne ocellata</i> Strömfelt	IIB
<i>Microspongium globosum</i> Reinke	IIB
<i>Mikrosyphar polysiphoniae</i> Kuckuck	IIB
<i>Myriactula arabica</i> (Kützing) Feldmann	IIB
<i>Myriactula rivulariae</i> (Suhr) Feldmann	IIB
<i>Myrionema orbiculare</i> J. Agardh	IIB
<i>Myrionema strangulans</i> Greville	IIB
<i>Myriotrichia clavaeformis</i> Harvey	IIB
<i>Nemacystus flexuosus</i> (C. Agardh) Kylin var. <i>giraudyi</i> (J. Agardh) de Jong	IIB
<i>Neosiphonia elongella</i> (Harvey) M.S. Kim & I.K.Lee	IIB
<i>Neosiphonia sertularioides</i> (Grateloup) K.W.Nam & P.J.Kang	IIB
<i>Nitophyllum punctatum</i> (Stackhouse) Greville	IIA
<i>Osmundea pelagiensis</i> G.Furnari	IIA
<i>Osmundea pinnatifida</i> (Hudson) Stackhouse	IIA
<i>Osmundea spectabilis</i> (Postels & Ruprecht) K.W.Nam	IIA
<i>Padina pavonica</i> (L.) Thivy	IB
<i>Palisada patentiramea</i> (Mont.) Cassano, Senties, Gil-Rodríguez & M.T. Fujii	IIA
<i>Palisada perforata</i> (Bory) K.W.Nam	IIA
<i>Percursaria percursa</i> (C.Agardh) Rosenvinge	IIB
<i>Petalonia fascia</i> (O.F. Müller) Kuntze	IIB
<i>Petalonia zosterifolia</i> (Reinke) Kuntze	IIB
<i>Peyssonnelia dubyi</i> P.Crouan & H.Crouan	IC
<i>Peyssonnelia squamaria</i> (S.G. Gmelin) Decaisne	IC
<i>Phaeophila dendroides</i> (P.L. Crouan et H.M. Crouan) Batters	IIB
<i>Phyllophora crispa</i> (Huds.) P.S.Dixon	IIA
<i>Phymatolithon calcareum</i> (Pallas) W.H.Adey & D.L.McKibbin	IC
<i>Phymatolithon lenormandii</i> (Aresch.) W.H.Adey	IC
<i>Pneophyllum confervicola</i> (Kützing) Y.M.Chamberlain	IC
<i>Pneophyllum fragile</i> Kützing	IC
<i>Polysiphonia atra</i> Zanardini	IIB
<i>Polysiphonia breviarticulata</i> (C.Agardh) Zanardini	IIB
<i>Polysiphonia brodiei</i> (Dillwyn) Sprengel	IIB
<i>Polysiphonia denudata</i> (Dillwyn) Greville ex Harvey	IIB

Appendix 1. Continued

<i>Polysiphonia deusta</i> (Roth) Sprengel	IIB
<i>Polysiphonia elongata</i> (Hudson) Sprengel	IIB
<i>Polysiphonia kampsaxii</i> Børgesen	IIB
<i>Polysiphonia morrowii</i> Harvey	IIB
<i>Polysiphonia nodulosa</i> J.Agardh	IIB
<i>Polysiphonia opaca</i> (C. Agardh) Moris <i>et</i> De Notaris	IIB
<i>Polysiphonia sanguinea</i> (C.Agardh) Zanardini	IIB
<i>Polysiphonia scopulorum</i> Harvey	IIB
<i>Porphyra umbilicalis</i> Kützing	IIB
<i>Protectocarpus speciosus</i> (Børgesen) Kornmann	IIB
<i>Pseudochlorodesmis furcellata</i> (Zanardini) Børgesen	IIB
<i>Pseudolithoderma adriaticum</i> (Hauck) Verlaque	IC
<i>Pterocladia capillacea</i> (S.G.Gmelin) Santelices & Hommersand	IIA
<i>Punctaria latifolia</i> Greville	IIB
<i>Punctaria plantaginea</i> (Roth) Greville	IIB
<i>Punctaria tenuissima</i> (C. Agardh) Greville	IIB
<i>Pyropia leucosticta</i> (Thuret) Neefus <i>et</i> J.Brodie	IIB
<i>Ralfsia verrucosa</i> (Areschoug) Areschoug	IC
<i>Rhizoclonium riparium</i> (Roth) Harvey	IIB
<i>Rhodochorton purpureum</i> (Lightfoot) Rosenvinge	IIB
<i>Rhodophyllis divaricata</i> (Stackhouse) Papenfuss	IIA
<i>Rhodymenia ardissoni</i> (Kuntze) Feldmann	IIA
<i>Rytiphlaea tinctoria</i> (Clemente) C.Agardh	IIA
<i>Sahlingia subintegra</i> (Rosenvinge) Kornmann	IC
<i>Sargassum vulgare</i> C.Agardh	IB
<i>Sauvageaugloia divaricata</i> (Clemente) Cremades	IIB
<i>Scytosiphon dotyi</i> M.J. Wynne	IIB
<i>Scytosiphon lomentaria</i> (Lyngbye) Link	IIB
<i>Spermatocnus paradoxus</i> (Roth) Kützing	IIB
<i>Spermothamnion repens</i> (Dillwyn) Rosenvinge	IIB
<i>Sphacelaria cirrosa</i> (Roth) C. Agardh	IIA
<i>Sphacelaria rigidula</i> Kützing	IIA
<i>Sphacelaria tribuloides</i> Meneghini	IIA
<i>Spongonema tomentosum</i> (Hudson) Kützing	IIB
<i>Spyridia filamentosa</i> (Wulfen) Harvey	IIB
<i>Stictyosiphon adriaticus</i> Kützing	IIB
<i>Stictyosiphon soriferus</i> (Reinke) Rosenvinge	IIB
<i>Stilophora nodulosa</i> (C. Agardh) P.C. Silva	IIB
<i>Stilophora tenella</i> (Esper) P.C. Silva	IIB
<i>Streblonema parasiticum</i> (Sauvageau) De Toni	IIB
<i>Striaria attenuata</i> (C. Agardh) Greville	IIB
<i>Stromatella monostromatica</i> (P.Dangeard) Kornmann & Sahling	IIB
<i>Stylonema alsidii</i> (Zanardini) K.M. Drew	IIB
<i>Stylonema cornu-cervi</i> Reinsch	IIB
<i>Trailiella intricata</i> Batters (tetrasporophyte of <i>Bonnemaisonia hamifera</i> Hariot)	IIB
<i>Ulothrix implexa</i> (Kützing) Kützing	IIB
<i>Ulva clathrata</i> (Roth) C.Agardh	IIB
<i>Ulva compressa</i> L.	IIB
<i>Ulva intestinalis</i> L.	IIB
<i>Ulva linza</i> L.	IIB
<i>Ulva polyclada</i> Kraft	IIB
<i>Ulva prolifera</i> O.F.Müller	IIB
<i>Ulva rigida</i> C.Agardh	IIB
<i>Ulva taeniata</i> (Setchell) Setchell & N.L.Gardner	IIB

Appendix 1. Continued

<i>Ulvella lens</i> P.L.Crouan <i>et</i> H.M.Crouan	IIB
<i>Ulvella leptochaete</i> (Huber) R.Nielsen, C.J.O'Kelly & B.Wysor	IIB
<i>Ulvella scutata</i> (Reinke) R.Nielsen	IIB
<i>Ulvella viridis</i> (Reinke) R.Nielsen, C.J.O'Kelly & B.Wysor	IIB
<i>Umbraulva dangeardii</i> M.J. Wynne & G.Furnari	IIB
<i>Vertebrata fucoides</i> (Hudson) Kuntze	IIB
<i>Vertebrata subulifera</i> (C.Agardh) Kuntze	IIB
<i>Vertebrata tripinnata</i> (Harvey) Kuntze	IIB
<hr/> ANGIOSPERMS <hr/>	
<i>Cymodocea nodosa</i> (Ucria) Ascherson	IB
<i>Posidonia oceanica</i> (L.) Delile	IA
<i>Zostera noltii</i> Hornemann	IB
